

Therapeutic targets of ataciguat

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Abstract

The endothelium regulates vascular and cardiac function by the release of endothelium-derived vasoactive compounds, the most important of which is nitric oxide (NO), which activates soluble guanylyl cyclase (sGC), inducing the formation of the second messenger cyclic guanosine monophosphate (cGMP). cGMP induces vasorelaxation, inhibits smooth muscle cell migration and proliferation, and tonically inhibits platelet activation, thereby preventing excessive aggregate formation and thrombosis. Furthermore, cGMP modulates myocardial oxygen metabolism, contractile function, ventricular hypertrophy and apoptosis, as well as fibrosis in the heart. Several cardiovascular diseases, such as congestive heart failure (CHF), coronary artery disease (CAD), hypertension, hypercholesterolemia, diabetes and early stages of atherosclerosis, are associated with a phenomenon commonly referred to as endothelial dysfunction. This is characterized by a significant reduction in NO bioavailability due to decreased formation or enhanced degradation. The common result is attenuated stimulation of sGC. In addition to impaired NO bioavailability, sGC itself can become dysfunctional, *e.g.*, under conditions of increased oxidative stress. Ataciguat, formerly referred to as HMR-1766, a novel anthranilic acid derivative, belongs to a new structural class of sGC activators capable of activating the oxidized form of sGC. Ataciguat has been shown to improve endothelial function and to reduce platelet activation in experimental diabetes. Chronic regular treatment with sGC activators such as ataciguat could therefore eventually constitute an effective therapy targeting deficient NO/cGMP signaling in hypertension, peripheral and coronary artery disease, heart failure, thrombosis and erectile dysfunction.

Endothelial function and nitric oxide-mediated guanylyl cyclase activation

The endothelium plays a crucial role in the control of vascular tone by releasing endothelium-derived autoids, the most important of which is nitric oxide (NO) (1). The necessity of intact endothelium as the source of the "endothelium-derived relaxing factor" (EDRF) was discovered in 1980 by Furchgott and Zawadzki (2). Seven years later, Moncada and Palmer demonstrated that EDRF is NO generated by an endothelial enzyme, endothelial NO synthase (eNOS), which converts the amino acid arginine to citrulline either under resting conditions or after stimulation (3).

NO is a ubiquitous, cell-permeable intercellular messenger which diffuses through the cell membrane to the underlying smooth muscle cells (SMCs). There it interacts with its specific molecular target, soluble guanylyl cyclase (sGC), which partially associates with the plasma membrane in a state of enhanced NO sensitivity (see Fig. 1) (4).

The mammalian NO-sensitive sGC is a heterodimeric heme protein existing in two isoforms with similar enzymatic properties (5). Binding of NO to the ferrous heme of sGC rapidly catalyzes the conversion of guanosine triphosphate (GTP) to cyclic guanosine monophosphate (cGMP). sGC heme iron has to be in the ferrous (Fe^{2+}) state for activation by NO. Upon NO binding, the iron is slightly moved out of the porphyrin plane, which is considered to trigger subsequent intramolecular rearrangements influencing the catalytic center (6).

The NO-induced cGMP signal exerts its effects through several intracellular mechanisms: cGMP-dependent protein kinases (cGKs), cGMP-regulated phosphodiesterases (PDEs) and cGMP-gated ion channels (5). cGK activation is the most prominent effector of NO in the cardiovascular system, controlling smooth muscle relaxation (7-9) and inhibition of platelet activation (10-16).

A common cGK substrate in SMCs and platelets is the vasodilator-stimulated phosphoprotein (VASP) (17-20). NO/cGMP-dependent phosphorylation of VASP plays a pivotal inhibitory role in the regulation of platelet activation (21). Phosphorylation of VASP correlates closely with inhibition of fibrinogen binding to platelet glycoprotein IIb/IIIa (gpIIb/IIIa) (11, 22). Increased cGMP induces phosphorylation of VASP preferentially at its serine residues 239 (Ser239) (21) and 157 (Ser157) (23), and

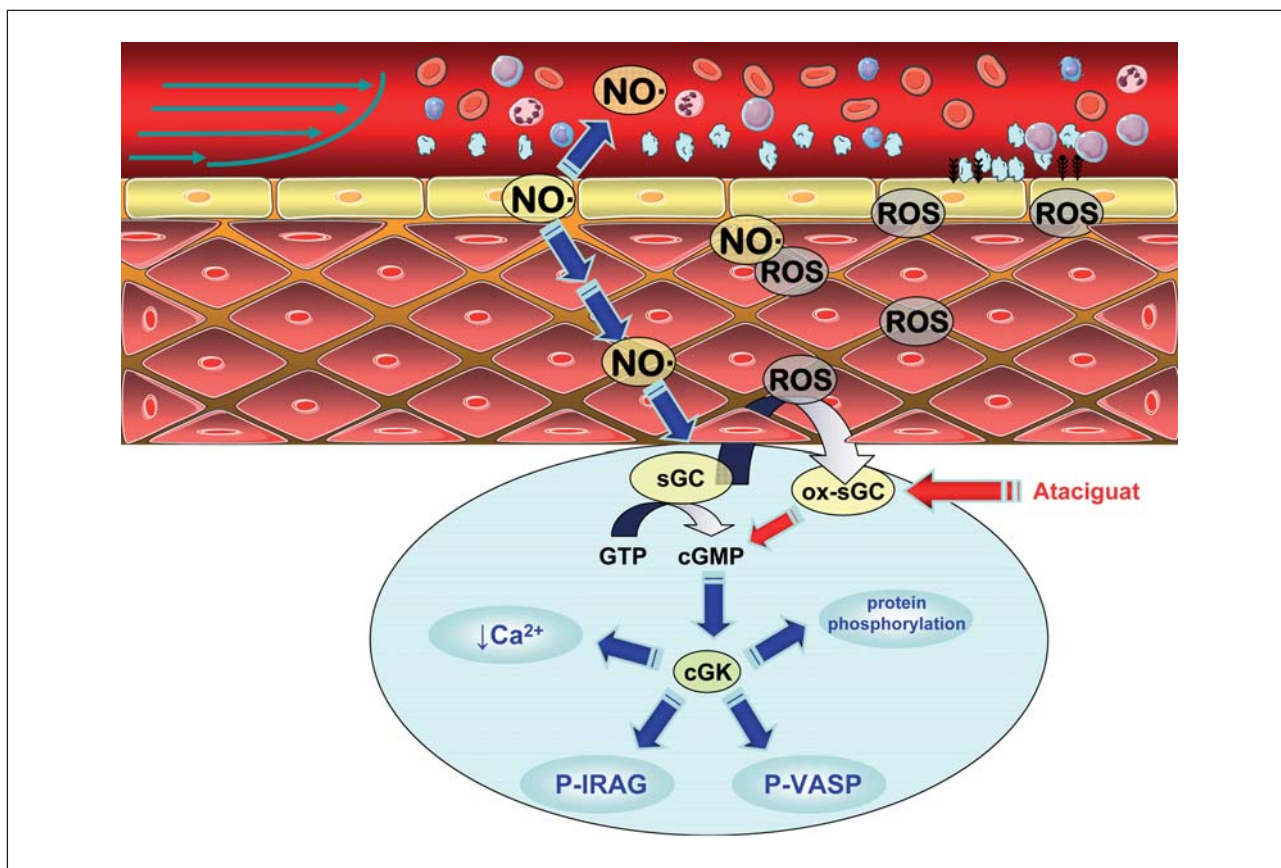


Fig. 1. Blood flow (green arrows) induces formation of nitric oxide (NO) in functional endothelial cells. Endothelium-derived NO exerts its effects within the vascular wall, targeting smooth muscle cells (SMCs) and on the luminal side interacts with blood cells, *e.g.*, it inhibits platelets and prevents their adhesion on the endothelium. In the target cell, NO activates NO-sensitive soluble guanylyl cyclase (sGC). Functional sGC catalyzes the reaction from guanosine triphosphate (GTP) to cyclic guanosine monophosphate (cGMP), which in turn activates the cGMP-dependent kinase (cGK). cGK ultimately reduces calcium influx (Ca^{2+}) and phosphorylates several substrate proteins, *e.g.*, the vasodilator-stimulated phosphoprotein (VASP) and the inositol-1,4,5-trisphosphate receptor I-associated protein (IRAG). Several cardiovascular diseases are associated with an increase in the generation of vascular reactive oxygen species (ROS) throughout the vessel wall. These scavenge NO and oxidize sGC (ox-sGC), such that the impaired remaining NO signal cannot activate sGC because ox-sGC is NO-insensitive. The novel direct sGC activator ataciguat is able to stimulate ox-sGC and therefore can improve cGMP-mediated activation of cGK in cardiovascular disease states with increased formation of ROS. See main text for details.

modulates platelet actin filament interactions (24). VASP phosphorylation affects initial sequences in platelet adhesion and activation (12, 25, 26). Using specific antibodies, VASP phosphorylation provides a sensitive monitor of defective NO/cGMP signaling, and reduced NO bioavailability in several pathophysiological states correlates with reduced VASP phosphorylation (27).

Endothelial dysfunction

Endothelial dysfunction was initially described as impaired vasodilatation in response to agonists such as acetylcholine and bradykinin, *e.g.*, in the forearm vasculature of patients with hypertension (28). Whereas the infusion of an NOS inhibitor blunts the vasodilator response to acetylcholine in control subjects, the arginine analogue does not significantly alter the response to

acetylcholine in hypertensive patients. This indicates that hypertensive patients have a defect in the endothelium-derived NO system that accounts for the impaired response to endothelium-dependent vasodilators (29). Compelling evidence suggests that endothelial dysfunction results from increased vascular production of superoxide anions in several pathophysiological states such as hypertension, heart failure and diabetes (30-33).

The renin-angiotensin system (RAS) is a central component of the physiological and pathological responses of the cardiovascular system. Its primary effector, angiotensin II, is implicated in inflammation, endothelial dysfunction, atherosclerosis, hypertension and congestive heart failure (CHF), in addition to its physiological vasoconstrictile and blood pressure-regulating effects (34). Inhibition of angiotensin-mediated signaling in patients with atherosclerosis reverses endothelial dys-

function by improving NO availability (35). Angiotensin II causes endothelial dysfunction and reduces vascular NO bioavailability by increasing vascular superoxide anion formation, enhanced vascular protein kinase C (PKC) activity and expression of the NADPH oxidase subunits nox1 , $\text{gp91}^{\text{phox}}$ and p22^{phox} (36). Because superoxide rapidly scavenges NO within the vascular wall, a reduction of bioactive NO might occur despite a compensatory increased NO generation.

Normal endothelial function plays a pivotal role in the suppression of SMC proliferation. NO and prostacyclin induce the formation of cGMP and cAMP, respectively. Both cyclic nucleotides regulate cell cycle molecules in human SMCs and suppress mitogenesis, proliferation and migration (37, 38). Chronic inhibition of NOS in rats changes SMC gene expression in favor of a shift towards cell proliferation (39). NO has antiproliferative properties and inhibits the angiotensin II-induced migration of SMCs (40). Gene transfer of cGK sensitizes cultured SMCs to the antiproliferative effects of NO/cGMP (41). Organic nitrates, which liberate NO, suppress proliferation and mitogenesis, an effect enhanced by PDE5 inhibition (42, 43). YC-1, an NO-independent activator of sGC (44), exerts vascular protection through inhibition of SMC proliferation (45).

In addition to these intramural effects of deficient NO signaling, impaired luminal NO bioavailability precipitates arterial thrombosis in animal models and in individuals with endothelial dysfunction (46). Chronic inhibition of NO formation in animal models is associated with impaired fibrinolysis, enhanced thrombin and tissue factor generation (47). Platelet activation is increased in disease states with impaired NO bioavailability, such as acute coronary syndromes (48), heart failure (49, 50), diabetes (51-53) and hypercholesterolemia (54). In patients with advanced atherosclerosis, an impaired endothelium-dependent release of NO leads to reduced platelet cGMP formation (55).

We previously demonstrated that acute reduction of NO bioavailability *in vivo* rapidly increases platelet activation in humans (16) and mice (52), which is immediately reversed by exogenous NO. This direct relationship between NO bioavailability and human platelet function *in vivo* suggests that platelet activation in healthy individuals is suppressed by tonic NO production, resulting in immediate platelet activation when NO production is inhibited (16).

The relevance of the NO/cGMP signaling pathway for platelet inhibition was demonstrated by several *in vitro* and *in vivo* studies: genetic deletion of the sGC $\beta 1$ subunit completely prevented NO-mediated inhibition of platelet aggregation (56), and platelet cGK proved essential to prevent platelet-endothelium adhesion and platelet-platelet aggregation after ischemia (25). Similarly, deletion of VASP increased platelet adhesion to the endothelium of ApoE^{-/-} mice and to the subendothelial matrix following endothelial denudation, which could not be rescued by exogenous NO (26). These results strengthen the central functional role for NO/cGMP/

cGK/VASP signaling in tonic platelet inhibition. The significance of disruption of this pathway for enhanced platelet activation in endothelial dysfunction is underlined by several studies, which demonstrate reduced platelet activation following positive pharmacological modulation of endothelial function: angiotensin or aldosterone blockade (49, 57), HMG-CoA reductase inhibition (50, 58) or direct stimulation of sGC with ataciguat (HMR-1766; sanofi-aventis) (53) reduced platelet activation and enhanced platelet VASP phosphorylation parallel to improved endothelial function in experimental CHF and diabetes.

Nitrate tolerance and eNOS uncoupling

The logical method of treating reduced NO bioavailability would be the supplementation of exogenous NO using NO donors such as organic nitrates. While this is a useful short-term approach during acute states of deficiency, prolonged nitrate treatment exerts several unfavorable effects, *e.g.*, it induces tolerance to nitrates and cross-tolerance to nitrovasodilators and endothelium-derived NO (59). Long-term treatment leads to enhanced superoxide anion production, at least partially facilitated by dysfunctional eNOS itself contributing to the so-called eNOS uncoupling (60), a phenomenon observed in several cardiovascular disease states, where eNOS generates superoxide instead of NO and enhanced eNOS activity might be deleterious (33, 61-63). In addition to scavenging of NO by superoxide, the reaction product peroxynitrite can oxidize the eNOS cofactor tetrahydrobiopterin, leading to uncoupling of eNOS. Restoration of endothelial tetrahydrobiopterin levels by transgenic approaches in genetic models of atherosclerosis reduces superoxide generation, normalizes endothelial function and attenuates disease progression (62). Similarly, normalization of tetrahydrobiopterin is generally able to restore eNOS-mediated NO formation and endothelial function in hypertension, hypercholesterolemia and diabetes (64-66). eNOS uncoupling and endothelial dysfunction are apparent in experimental diabetes and in diabetic patients (33, 67) despite the fact that eNOS expression is actually increased. Therefore, approaches merely aimed at enhanced eNOS activity would not improve NO bioavailability, but would aggravate endothelial dysfunction.

sGC dysfunction

In principal, two therapeutic approaches remain to enhance defective cGMP signaling: 1) preventing the degradation of the remaining cGMP by PDE inhibitors; or 2) increased stimulation of sGC by direct activators of the enzyme.

sGC heme iron has to be in the ferrous (Fe^{2+}) state for activation by NO. While NO increases sGC activity, reactive oxygen species (ROS) exert opposite effects on the enzyme and functionally counteract NO (68). Activation by NO is lost but basal activity is preserved if the heme

iron is oxidized to the ferric (Fe^{3+}) state (69). Increased oxidative stress decreases the expression and impairs NO-induced activation of heme-containing sGC, making vasodilator therapy with NO donors less effective. Under conditions of enhanced oxidative stress *in vivo* in several disease states, including diabetes, sGC is indistinguishable from the *in vitro* oxidized/heme-free enzyme (69). The oxidized/heme-free sGC variant is unresponsive to NO and prone to degradation.

Thus, trying to enhance cGMP signaling by PDE inhibition is hampered by the dependence on a sufficiently high remaining cGMP signal. As described above, this signal is strongly affected by reduced formation of NO, uncoupling of eNOS, enhanced scavenging of NO by ROS and reduced sensitivity of sGC for NO. Therefore, heme-dependent and -independent activators of sGC have recently emerged as potential modulators of defective NO/cGMP-mediated signaling (70). Novel NO-independent sGC activators such as Bay-58-2667 or ataciguat stabilize and/or activate the NO-insensitive sGC variant (69, 71). These drugs may represent a useful therapeutic approach to preferentially dilate diseased blood vessels.

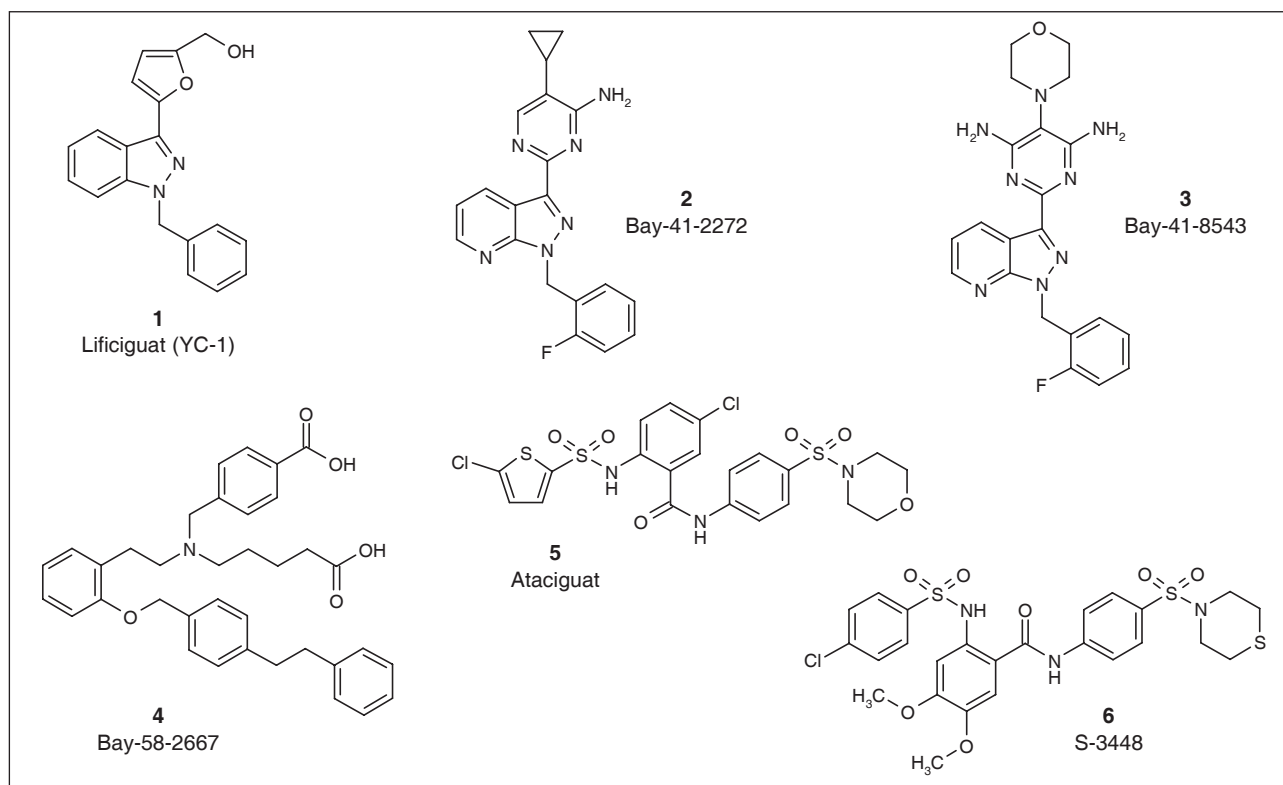
sGC activators

The first substance characterized as an NO-independent, heme-dependent stimulator of sGC was YC-1 (lifiguat, **1**) (72). YC-1 potentiates the efficacy of nitrovasodilators (44) and causes persistent elevation of intravascular cGMP levels *in vivo* by activating sGC and

inhibiting cGMP breakdown (73). Other heme-dependent sGC stimulators were synthesized based on YC-1, including Bay-41-2272 (**2**) and Bay-41-8543 (**3**), which are both approximately two orders of magnitude more potent than YC-1 (70).

Activation of sGC with Bay-41-2272 reverses hemodynamic and structural changes associated with monocrotaline- and chronic hypoxia-induced experimental pulmonary hypertension. This effect is partially dependent on endogenous NO generated by eNOS (74). Chronic sGC activation in mice inhibited endothelial P-selectin expression and leukocyte recruitment in mesenteric postcapillary venules following an inflammatory response elicited by IL-1 β (75). NO and prostacyclin exhibit synergistic activity with Bay-41-2272 to attenuate platelet aggregation and significantly reduce blood pressure (76). Chronic sGC activation avoided the hypertension induced by systemic NOS inhibition in rats and prevented cardiac hypertrophy and fibrosis (77).

Two sGC activators, Bay-58-2667 (**4**) and Bay-41-2272, have been investigated in pacing-induced heart failure. Acute intravenous sGC activation in dogs potentially unloaded the heart, increased cardiac output and renal blood flow, and preserved renal function without further neurohumoral activation (78, 79). Because the oxidized/heme-free sGC variant is unresponsive to NO and prone to degradation, novel NO-independent sGC activators such as Bay-58-2667, which stabilize and activate the NO-insensitive sGC variant, might represent a useful therapeutic approach, preferentially dilating diseased blood vessels (69).



Ataciguat

Ataciguat (**5**) is a novel anthranilic acid derivative activating heme-oxidized sGC (71). Ataciguat and S-3448 (**6**) comprise a new structural class of sGC activators capable of activating the oxidized and/or heme-free forms of sGC (71). The potential interesting application of this class of substances is indicated by the efficacy in initial pharmacological studies: both compounds increased cGMP levels in cultured rat aortic SMCs and relaxed isolated endothelium-denuded rat aorta (71). Similarly, chronic treatment with ataciguat improved endothelial function and reversed the change to a noncontractile phenotype in aortae from diabetic rats (80). In ApoE^{-/-} mice, ataciguat improved endothelium-dependent vasorelaxation and reduced atherosclerosis (81).

As observed with other sGC activators, ataciguat attenuated the development of pulmonary hypertension and right heart hypertrophy, preserved systemic arterial blood pressure, improved gas exchange and cardiac output in monocrotaline-induced pulmonary hypertension (82). In a model of transforming growth factor- β (TGF- β)-induced cardiac fibroblast extracellular matrix synthesis, ataciguat inhibited fibronectin protein synthesis in a concentration-dependent manner and prevented the differentiation of fibroblasts into myofibroblasts (83).

Pharmacological sGC activation exerts antiplatelet properties *in vitro* by increasing platelet cGMP, the main mediator of NO effects in platelets (13, 76, 84). Ataciguat elicited phosphorylation of the cGK substrate VASP at Ser239 and reduced platelet reactivity (53). The results from *in vitro* and acute *in vivo* experiments in diabetic rats suggest that the stimulation of platelet sGC by ataciguat was independent of sGC dysfunction (53). Chronic sGC stimulation *in vivo* attenuated the activation of gpIIb/IIIa (determined by platelet fibrinogen binding), as well as platelet degranulation (determined by P-selectin surface expression). P-selectin can participate in platelet adhesion to the endothelium and is responsible for platelet-leukocyte adhesion (85, 86). P-selectin-expressing platelets play a pivotal role in the interaction of activated platelets with leukocytes and the exacerbation of atherosclerosis (87). Earlier studies reported inhibition of P-selectin and CD40 ligand surface expression by cAMP/cGMP-dependent protein kinases (88), the activation of which can be monitored by VASP phosphorylation (23).

Patients with diabetes have an increased risk of thrombosis and accelerated atherogenesis. Platelet degranulation in patients with diabetes is associated with progression of proatherosclerotic vessel wall modification (89). Ataciguat induces vasorelaxation of porcine coronary arteries (71) and might therefore—together with inhibition of platelet activation—exert potential beneficial effects in stable and unstable CAD. Activation of oxidized sGC by ataciguat might be useful to overcome impaired platelet NO responsiveness, which in addition to decreased endothelial formation of NO is a predictor of increased mortality and cardiovascular morbidity in patients with acute coronary syndromes (90).

Deficient endothelium-dependent and -independent vasodilatation occurs often before the development of other overt functional or structural systemic vascular disease in patients with erectile dysfunction (91). Erectile dysfunction in diabetic men correlates with endothelial dysfunction, and reduced NO bioavailability/activity might provide a unifying explanation (92). Similar to PDE5 inhibitors, which prevent the degradation of cGMP, ataciguat sufficiently relaxes human corpus cavernosum (71).

Currently, the ACCELA study (ClinicalTrials.gov Identifier: NCT00443287) is the first clinical trial assessing the efficacy and safety of ataciguat in patients with peripheral arterial occlusive disease (PAOD). The primary objective is to investigate whether a 26-week treatment with ataciguat (three different doses) in addition to clopidogrel results in an improvement in walking capacity in patients suffering from intermittent claudication due to Fontaine stage II peripheral arterial disease, compared to placebo. The effects will be compared with cilostazol, which reversibly inhibits platelets via selective antagonism of PDE3.

Conclusions

Chronic regular treatment with sGC activators such as ataciguat may constitute an effective therapy targeting hypertension, peripheral and coronary artery disease, as well as improving cardiac function, inhibiting platelet activation and thrombosis, and preventing erectile dysfunction. sGC activation with ataciguat combines enhancement of cGMP signaling independent of NO bioavailability. Given its ability to activate the heme-oxidized NO-insensitive form of sGC, ataciguat might still be active under certain conditions of oxidative stress, when other sGC activators or NO itself are ineffective because of sGC dysfunction. Ataciguat might therefore represent a useful treatment approach in atherosclerotic cardiovascular disease.

References

1. Busse, R., Fleming, I. *Regulation of endothelium-derived vasoactive autacoid production by hemodynamic forces*. Trends Pharmacol Sci 2003, 24: 24-9.
2. Furchgott, R.F., Zawadzki, J.V. *The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine*. Nature 1980, 288: 373-6.
3. Palmer, R.M.J., Ferrige, A.G., Moncada, S. *Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor*. Nature 1987, 327: 524-6.
4. Zabel, U., Kleinschnitz, C., Oh, P. et al. *Calcium-dependent membrane association sensitizes soluble guanylyl cyclase to nitric oxide*. Nat Cell Biol 2002, 4: 307-11.
5. Friebe, A., Koesling, D. *Regulation of nitric oxide-sensitive guanylyl cyclase*. Circ Res 2003, 93: 96-105.
6. Wedel, B., Humbert, P., Harteneck, C. et al. *Mutation of His-105 in the beta 1 subunit yields a nitric oxide-insensitive form of*

- soluble guanylyl cyclase. *Proc Natl Acad Sci USA* 1994, 91: 2592-6.
7. Sausbier, M., Schubert, R., Voigt, V. et al. *Mechanisms of NO/cGMP-dependent vasorelaxation*. *Circ Res* 2000, 87: 825-30.
 8. Moncada, S., Higgs, A. *Mechanisms of disease - The L-arginine/nitric oxide pathway*. *New Eng J Med* 1993, 329: 2002-12.
 9. Pfeifer, A., Klatt, P., Massberg, S. et al. *Defective smooth muscle regulation in cGMP kinase I-deficient mice*. *EMBO J* 1998, 17: 3045-51.
 10. Radomski, M.W., Palmer, R.M., Moncada, S. *An L-arginine/nitric oxide pathway present in human platelets regulates aggregation*. *Proc Natl Acad Sci USA* 1990, 87: 5193-7.
 11. Hauser, W., Knobloch, K.P., Eigenthaler, M. et al. *Megakaryocyte hyperplasia and enhanced agonist-induced platelet activation in vasodilator-stimulated phosphoprotein knockout mice*. *Proc Natl Acad Sci USA* 1999, 96: 8120-5.
 12. Schwarz, U.R., Walter, U., Eigenthaler, M. *Taming platelets with cyclic nucleotides*. *Biochem Pharmacol* 2001, 62: 1153-61.
 13. Moro, M.A., Russel, R.J., Cellek, S. et al. *cGMP mediates the vascular and platelet actions of nitric oxide: Confirmation using an inhibitor of the soluble guanylyl cyclase*. *Proc Natl Acad Sci USA* 1996, 93: 1480-5.
 14. de Belder, A.J., MacAllister, R., Radomski, M.W., Moncada, S., Vallance, P.J. *Effects of S-nitroso-glutathione in the human forearm circulation: Evidence for selective inhibition of platelet activation*. *Cardiovasc Res* 1994, 28: 691-4.
 15. Alheid, U., Frolich, J.C., Forstermann, U. *Endothelium-derived relaxing factor from cultured human endothelial cells inhibits aggregation of human platelets*. *Thromb Res* 1987, 47: 561-71.
 16. Schäfer, A., Wiesmann, F., Neubauer, S., Eigenthaler, M., Bauersachs, J., Channon, K.M. *Rapid regulation of platelet activation in vivo by nitric oxide*. *Circulation* 2004, 109: 1819-22.
 17. Aszódi, A., Pfeifer, A., Ahmad, M. et al. *The vasodilator-stimulated phosphoprotein (VASP) is involved in cGMP- and cAMP-mediated inhibition of agonist-induced platelet aggregation, but is dispensable for smooth muscle function*. *EMBO J* 1999, 18: 37-48.
 18. Butt, E., Abel, K., Krieger, M. et al. *cAMP- and cGMP-dependent protein kinase phosphorylation sites of the focal adhesion vasodilator-stimulated phosphoprotein (VASP) in vitro and in intact human platelets*. *J Biol Chem* 1994, 269: 14509-17.
 19. Schwarz, U.R., Geiger, J., Walter, U., Eigenthaler, M. *Flow cytometry analysis of intracellular VASP phosphorylation for the assessment of activating and inhibitory signal transduction pathways in human platelets*. *Thromb Haemost* 1999, 82: 1145-52.
 20. Walter, U., Eigenthaler, M., Geiger, J., Reinhard, M. *Role of cyclic nucleotide-dependent protein kinases and their common substrate VASP in the regulation of human platelets*. *Adv Exp Med Biol* 1993, 344: 237-49.
 21. Eigenthaler, M., Nolte, C., Halbruegge, M., Walter, U. *Concentration and regulation of cyclic nucleotides, cyclic-nucleotide-dependent protein kinases and one of their major substrates in human platelets: Estimating the rate of cAMP-regulated and cGMP-regulated protein phosphorylation in intact cells*. *Eur J Biochem* 1992, 205: 471-81.
 22. Horstrup, K., Jablonka, B., Honig-Liedl, P., Just, M., Kochsiek, K., Walter, U. *Phosphorylation of focal adhesion vasodilator-stimulated phosphoprotein at Ser157 in intact human platelets correlates with fibrinogen receptor inhibition*. *Eur J Biochem* 1994, 225: 21-7.
 23. Schäfer, A., Vollkommer, T., Burkhardt, M. et al. *Endothelium-dependent and -independent relaxation and VASP serines 157/239 phosphorylation by cyclic nucleotide-elevating vasodilators in rat aorta*. *Biochem Pharmacol* 2003, 65: 397-405.
 24. Bearer, E.L., Prakash, J.M., Manchester, R.D., Allen, P.G. *VASP protects actin filaments from gelsolin: An in vitro study with implications for platelet actin reorganizations*. *Cell Motil Cytoskeleton* 2000, 47: 351-64.
 25. Massberg, S., Sausbier, M., Klatt, P. et al. *Increased adhesion and aggregation of platelets lacking cyclic guanosine 3',5'-monophosphate kinase I*. *J Exp Med* 1999, 189: 1255-64.
 26. Massberg, S., Grüner, S., Konrad, I. et al. *Enhanced in vivo platelet adhesion in vasodilator-stimulated phosphoprotein (VASP)-deficient mice*. *Blood* 2004, 103: 136-42.
 27. Oelze, M., Mollnau, H., Hoffmann, N. et al. *Vasodilator-stimulated phosphoprotein serine 239 phosphorylation as a sensitive monitor of defective nitric oxide/cGMP signaling and endothelial dysfunction*. *Circ Res* 2000, 87: 999-1005.
 28. Panza, J.A., Quyyumi, A.A., Brush, J.E. Jr., Epstein, S.E. *Abnormal endothelium-dependent vascular relaxation in patients with essential hypertension*. *New Eng J Med* 1990, 323: 22-7.
 29. Panza, J.A., Casino, P.R., Kilcoyne, C.M., Quyyumi, A.A. *Role of endothelium-derived nitric oxide in the abnormal endothelium-dependent vascular relaxation of patients with essential hypertension*. *Circulation* 1993, 87: 1468-74.
 30. Bauersachs, J., Bouloumié, A., Mülsch, A., Wiemer, G., Fleming, I., Busse, R. *Vasodilator dysfunction in aged spontaneously hypertensive rats: Changes in NO synthase III and soluble guanylyl cyclase expression, and in superoxide anion production*. *Cardiovasc Res* 1998, 37: 772-9.
 31. Bauersachs, J., Bouloumié, A., Fraccarollo, D., Hu, K., Busse, R., Ertl, G. *Endothelial dysfunction in chronic myocardial infarction despite increased vascular endothelial nitric oxide synthase and soluble guanylyl cyclase expression: Role of enhanced vascular superoxide production*. *Circulation* 1999, 100: 292-8.
 32. Bauersachs, J., Schäfer, A. *Endothelial dysfunction in heart failure: Mechanisms and therapeutic approaches*. *Curr Vasc Pharmacol* 2004, 2: 115-24.
 33. Hink, U., Li, H., Mollnau, H. et al. *Mechanisms underlying endothelial dysfunction in diabetes mellitus*. *Circ Res* 2001, 88: e14-22.
 34. Mehta, P.K., Griending, K.K. *Angiotensin II cell signaling: Physiological and pathological effects in the cardiovascular system*. *Am J Physiol Cell Physiol* 2007, 292: C82-97.
 35. Prasad, A., Tupas-Habib, T., Schenke, W.H. et al. *Acute and chronic angiotensin-1 receptor antagonism reverses endothelial dysfunction in atherosclerosis*. *Circulation* 2000, 101: 2349-54.
 36. Mollnau, H., Wendt, M., Szocs, K. et al. *Effects of angiotensin II infusion on the expression and function of NAD(P)H oxidase and components of nitric oxide/cGMP signaling*. *Circ Res* 2002, 90: E58-65.

37. Fukumoto, S., Koyama, H., Hosoi, M. et al. *Distinct role of cAMP and cGMP in the cell cycle control of vascular smooth muscle cells: cGMP delays cell cycle transition through suppression of cyclin D1 and cyclin-dependent kinase 4 activation.* Circ Res 1999, 85: 985-91.
38. Rybalkin, S.D., Yan, C., Bornfeldt, K.E., Beavo, J.A. *Cyclic GMP phosphodiesterases and regulation of smooth muscle function.* Circ Res 2003, 93: 280-91.
39. Dupuis, M., Soubrier, F., Brocheriou, I. et al. *Profiling of aortic smooth muscle cell gene expression in response to chronic inhibition of nitric oxide synthase in rats.* Circulation 2004, 110: 867-73.
40. Dubey, R.K., Jackson, E.K., Lüscher, T.F. *Nitric oxide inhibits angiotensin II-induced migration of rat aortic smooth muscle cell: Role of cyclic-nucleotides and angiotensin1 receptors.* J Clin Invest 1995, 96: 141-9.
41. Chiche, J.-D., Schlutsmeier, S.M., Bloch, D.B. et al. *Adenovirus-mediated gene transfer of cGMP-dependent protein kinase increases the sensitivity of cultured vascular smooth muscle cells to the antiproliferative and pro-apoptotic effects of nitric oxide/cGMP.* J Biol Chem 1998, 273: 34263-71.
42. Osinski, M.T., Rauch, B.H., Schror, K. *Antimitogenic actions of organic nitrates are potentiated by sildenafil and mediated via activation of protein kinase A.* Mol Pharmacol 2001, 59: 1044-50.
43. Young, D.V., Serebryanik, D., Janero, D.R., Tam, S.W. *Suppression of proliferation of human coronary artery smooth muscle cells by the nitric oxide donor, S-nitrosoglutathione, is cGMP-independent.* Mol Cell Biol Res Comm 2000, 4: 32-6.
44. Mulsch, A., Bauersachs, J., Schäfer, A., Stasch, J.P., Kast, R., Busse, R. *Effect of YC-1, an NO-independent, superoxide-sensitive stimulator of soluble guanylyl cyclase, on smooth muscle responsiveness to nitrovasodilators.* Br J Pharmacol 1997, 120: 681-9.
45. Tulis, D.A., Bohl Masters, K.S., Lipke, E.A. et al. *YC-1-mediated vascular protection through inhibition of smooth muscle cell proliferation and platelet function.* Biochem Biophys Res Commun 2002, 291: 1014-21.
46. Loscalzo, J. *Nitric oxide insufficiency, platelet activation, and arterial thrombosis.* Circ Res 2001, 88: 756-62.
47. Corseaux, D., Ollivier, V., Fontaine, V. et al. *Hemostasis imbalance in experimental hypertension.* Mol Med 2002, 8: 169-78.
48. Heeschen, C., Dimmeler, S., Hamm, C.W. et al. *Soluble CD40 ligand in acute coronary syndromes.* New Eng J Med 2003, 348: 1104-11.
49. Schäfer, A., Fraccarollo, D., Hildemann, S. et al. *Inhibition of platelet activation in congestive heart failure by selective aldosterone receptor antagonism and ACE inhibition: Role of endothelial function and platelet VASP phosphorylation.* Thromb Haemost 2003, 89: 1024-30.
50. Schäfer, A., Fraccarollo, D., Eigenthaler, M. et al. *Rosuvastatin reduces platelet activation in heart failure: Role of nitric oxide bioavailability.* Arterioscler Thromb Vasc Biol 2005, 25: 1071-7.
51. Winocour, P.D. *Platelet abnormalities in diabetes mellitus.* Diabetes 1992, 41(Suppl. 2): 26-31.
52. Schäfer, A., Alp, N.J., Cai, S. et al. *Reduced vascular NO bioavailability in diabetes increases platelet activation in vivo.* Arterioscler Thromb Vasc Biol 2004, 24: 1720-6.
53. Schäfer, A., Flierl, U., Kobsar, A., Eigenthaler, M., Ertl, G., Bauersachs, J. *Soluble guanylyl cyclase activation with HMR 1766 attenuates platelet activation in diabetic rats.* Arterioscler Thromb Vasc Biol 2006, 26: 2813-8.
54. Nimpf, J., Wurm, H., Kostner, G.M., Kenner, T. *Platelet activation in normo- and hyperlipoproteinemias.* Basic Res Cardiol 1986, 81: 437-53.
55. Andrews, N.P., Husain, M., Dakak, N., Quyyumi, A.A. *Platelet inhibitory effect of nitric oxide in the human coronary circulation: Impact of endothelial dysfunction.* J Am Coll Cardiol 2001, 37: 510-6.
56. Friebe, A., Mergia, E., Dangel, O., Lange, A., Koesling, D. *Fatal gastrointestinal obstruction and hypertension in mice lacking nitric oxide-sensitive guanylyl cyclase.* Proc Natl Acad Sci USA 2007, 104: 7699-704.
57. Schäfer, A., Flierl, U., Vogt, C. et al. *Telmisartan improves vascular function and reduces platelet activation in rats with streptozotocin-induced diabetes mellitus.* Pharmacol Res 2007, In press.
58. Schäfer, A., Fraccarollo, D., Vogt, C. et al. *Improved endothelial function and reduced platelet activation by chronic HMG-CoA-reductase inhibition with rosuvastatin in rats with streptozotocin-induced diabetes mellitus.* Biochem Pharmacol 2007, 73: 1367-75.
59. Mülsch, A., Oelze, M., Kloss, S. et al. *Effects of in vivo nitroglycerin treatment on activity and expression of the guanylyl cyclase and cGMP-dependent protein kinase and their downstream target vasodilator-stimulated phosphoprotein in aorta.* Circulation 2001, 103: 2188-94.
60. Münzel, T., Li, H., Mollnau, H. et al. *Effects of long-term nitroglycerin treatment on endothelial nitric oxide synthase (NOS III) gene expression, NOS III-mediated superoxide production, and vascular NO bioavailability.* Circ Res 2000, 86: E7-12.
61. Alp, N.J., Mussa, S., Khoo, J. et al. *Tetrahydrobiopterin-dependent preservation of nitric oxide-mediated endothelial function in diabetes by targeted transgenic GTP-cyclohydrolase I over-expression.* J Clin Invest 2003, 112: 725-35.
62. Alp, N.J., McAteer, M.A., Khoo, J., Choudhury, R.P., Channon, K.M. *Increased endothelial tetrahydrobiopterin synthesis by targeted transgenic GTP-cyclohydrolase I overexpression reduces endothelial dysfunction and atherosclerosis in ApoE-knockout mice.* Arterioscler Thromb Vasc Biol 2004, 24: 445-50.
63. Laursen, J.B., Somers, M., Kurz, S. et al. *Endothelial regulation of vasomotion in apoE-deficient mice: Implications for interactions between peroxynitrite and tetrahydrobiopterin.* Circulation 2001, 103: 1282-8.
64. Alp, N.J., Channon, K.M. *Regulation of endothelial nitric oxide synthase by tetrahydrobiopterin in vascular disease.* Arterioscler Thromb Vasc Biol 2004, 24: 413-20.
65. Cosentino, F., Luscher, T.F. *Tetrahydrobiopterin and endothelial nitric oxide synthase activity.* Cardiovasc Res 1999, 43: 274-8.
66. Vazquez-Vivar, J., Martasek, P., Whitsett, J., Joseph, J., Kalyanaram, B. *The ratio between tetrahydrobiopterin and oxi-*

- dized tetrahydrobiopterin analogues controls superoxide release from endothelial nitric oxide synthase: An EPR spin trapping study. *Biochem J* 2002, 362: 733-9.
67. Guzik, T.J., Mussa, S., Gastaldi, D. et al. *Mechanisms of increased vascular superoxide production in human diabetes mellitus: Role of NAD(P)H oxidase and endothelial nitric oxide synthase*. *Circulation* 2002, 105: 1656-62.
68. Munzel, T., Daiber, A., Ullrich, V., Mulsch, A. *Vascular consequences of endothelial nitric oxide synthase uncoupling for the activity and expression of the soluble guanylyl cyclase and the cGMP-dependent protein kinase*. *Arterioscler Thromb Vasc Biol* 2005, 25: 1551-7.
69. Stasch, J.P., Schmidt, P.M., Nedvetsky, P.I. et al. *Targeting the heme-oxidized nitric oxide receptor for selective vasodilatation of diseased blood vessels*. *J Clin Invest* 2006, 116: 2552-61.
70. Evgenov, O.V., Pacher, P., Schmidt, P.M., Hasko, G., Schmidt, H.H., Stasch, J.P. *NO-independent stimulators and activators of soluble guanylate cyclase: Discovery and therapeutic potential*. *Nat Rev Drug Discov* 2006, 5: 755-68.
71. Schindler, U., Strobel, H., Schonafinger, K. et al. *Biochemistry and pharmacology of novel anthranilic acid derivatives activating heme-oxidized soluble guanylyl cyclase*. *Mol Pharmacol* 2006, 69: 1260-8.
72. Ko, F.N., Wu, C.C., Kuo, S.C., Lee, F.Y., Teng, C.M. *YC-1, a novel activator of platelet guanylate cyclase*. *Blood* 1994, 84: 4226-33.
73. Galle, J., Zabel, U., Hubner, U. et al. *Effects of the soluble guanylyl cyclase activator, YC-1, on vascular tone, cyclic GMP levels and phosphodiesterase activity*. *Br J Pharmacol* 1999, 127: 195-203.
74. Dumitrascu, R., Weissmann, N., Ghofrani, H.A. et al. *Activation of soluble guanylate cyclase reverses experimental pulmonary hypertension and vascular remodeling*. *Circulation* 2006, 113: 286-95.
75. Ahluwalia, A., Foster, P., Scotland, R.S. et al. *Antiinflammatory activity of soluble guanylate cyclase: cGMP-dependent down-regulation of P-selectin expression and leukocyte recruitment*. *Proc Natl Acad Sci USA* 2004, 101: 1386-91.
76. Hobbs, A.J., Moncada, S. *Antiplatelet properties of a novel, non-NO-based soluble guanylate cyclase activator, BAY 41-2272*. *Vascul Pharmacol* 2003, 40: 149-54.
77. Zanfolin, M., Faro, R., Araujo, E.G., Guaraldo, A.M., Antunes, E., De Nucci, G. *Protective effects of BAY 41-2272 (sGC stimulator) on hypertension, heart, and cardiomyocyte hypertrophy induced by chronic L-NAME treatment in rats*. *J Cardiovasc Pharmacol* 2006, 47: 391-5.
78. Boerrigter, G., Costello-Boerrigter, L.C., Cataliotti, A., Lapp, H., Stasch, J.P., Burnett, J.C. Jr. *Targeting heme-oxidized soluble guanylate cyclase in experimental heart failure*. *Hypertension* 2007, 49: 1128-33.
79. Boerrigter, G., Costello-Boerrigter, L.C., Cataliotti, A. et al. *Cardiorenal and humoral properties of a novel direct soluble guanylate cyclase stimulator BAY 41-2272 in experimental congestive heart failure*. *Circulation* 2003, 107: 686-9.
80. Schäfer, A., Vogt, C., Fraccarollo, D., Bauersachs, J. *Improvement of vascular dysfunction in diabetes by chronic guanylyl cyclase activation*. *Circulation* 2005, 112: II-301.
81. Wassmann, S., van Eickels, M., Czech, T., Oelze, M., Strobel, H., Nickenig, G. *Chronic activation of soluble guanylate cyclase improves endothelial function and reduces atherosclerosis in apolipoprotein E-deficient mice*. *Eur Heart J* 2006, 27(4S): Abst.
82. Klein, M., Vogelsberger, S., Schindler, U. et al. *Treatment with a nitric oxide independent activator of the soluble guanylyl cyclase dramatically improves monocrotaline-induced pulmonary hypertension*. *Z Kardiol Suppl* 2005, Abst V910.
83. Illiano, S.C., Riva, L., Bouloy, M., Beauverger, P., O'Connor, S.E. *Effect of HMR1766, a soluble guanylate cyclase activator, on differentiation of cardiac fibroblasts and extracellular matrix synthesis induced by TGFbeta*. *Circulation* 2006, 114: Abst S281.
84. Friebe, A., Mullershausen, F., Smolenski, A., Walter, U., Schultz, G., Koesling, D. *YC-1 potentiates nitric oxide- and carbon monoxide-induced cyclic GMP effects in human platelets*. *Mol Pharmacol* 1998, 54: 962-7.
85. Furie, B., Furie, B.C., Flaumenhaft, R. *A journey with platelet P-selectin: The molecular basis of granule secretion, signalling and cell adhesion*. *Thromb Haemost* 2001, 86: 214-21.
86. Li, N., Hu, H., Lindqvist, M., Wikstrom-Jonsson, E., Goodall, A.H., Hjerdahl, P. *Platelet-leukocyte cross talk in whole blood*. *Arterioscler Thromb Vasc Biol* 2000, 20: 2702-8.
87. Huo, Y., Schober, A., Forlow, S.B. et al. *Circulating activated platelets exacerbate atherosclerosis in mice deficient in apolipoprotein E*. *Nat Med* 2003, 9: 61-7.
88. Schwarz, U.R., Kobsar, A.L., Koksche, M., Walter, U., Eigenthaler, M. *Inhibition of agonist-induced p42 and p38 mitogen-activated protein kinase phosphorylation and CD40 ligand/P-selectin expression by cyclic nucleotide-regulated pathways in human platelets*. *Biochem Pharmacol* 2000, 60: 1399-407.
89. Fateh-Moghadam, S., Li, Z., Ersel, S. et al. *Platelet degranulation is associated with progression of intima-media thickness of the common carotid artery in patients with diabetes mellitus type II*. *Arterioscler Thromb Vasc Biol* 2005, 25: 1299-303.
90. Willoughby, S.R., Stewart, S., Holmes, A.S., Chirkov, Y.Y., Horowitz, J.D. *Platelet nitric oxide responsiveness: A novel prognostic marker in acute coronary syndromes*. *Arterioscler Thromb Vasc Biol* 2005, 25: 2661-6.
91. Kaiser, D.R., Billups, K., Mason, C., Wetterling, R., Lundberg, J.L., Bank, A.J. *Impaired brachial artery endothelium-dependent and -independent vasodilation in men with erectile dysfunction and no other clinical cardiovascular disease*. *J Am Coll Cardiol* 2004, 43: 179-84.
92. De Angelis, L., Marfella, M.A., Siniscalchi, M. et al. *Erectile and endothelial dysfunction in Type II diabetes: A possible link*. *Diabetologia* 2001, 44: 1155-60.